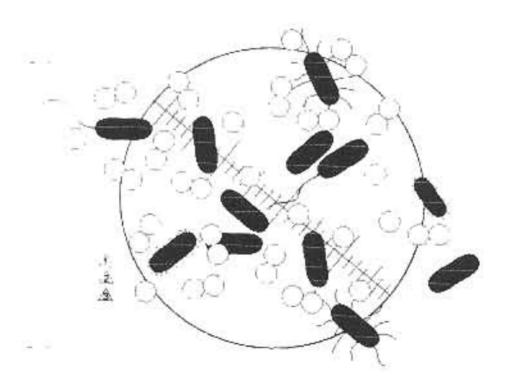
## Dept. of Science & Technology SEP 0 5 20 Grande Prairie Regional College

### MI 2650 General Microbiology

# Course Outline W 1999-2000



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Schedule:

Classes - Tuesday & Thursday 1300-1420 Labs - Friday 1200-1550

Description:

This course covers aspects of bacterial physiology such as nutrient uptake, metabolism, extracellular proteins, chemotaxis and differentiation. Symbiotic associations and interaction of microbes with the environment are major topics. Basic principles of industrial microbiology and the use of biotechnology for the production of economically and medically important substances will be covered. Laboratory exercises are designed to compliment the material included in the classes.

Text-book:

Brock - Biology of Microorganisms (9th edition)
MADIGAN, MARTINKO & PARKER (2000)
Prentice-Hall Publishers

This text-book is recommended for the course - it is not compulsory. For extra help with the text, Prentice-Hall Publishers are making available a web page containing Chapter summaries, self-tests, and other information that you may find useful. The URL address for this web-page is:

http://www.prenhall.com/~bookbind/pubbooks/brock

A list of relevant articles will be provided to each student, and it is strongly recommended that they be read. The recommended reading list is fairly long, therefore the most important articles will also be placed in a binder on reserve in the library.

The following books have also been placed on reserve in the GPRC Library, and students are advised to take advantage of their availability.

- ALCAMO, I.E. (1997) Fundamentals of Microbiology (5th Edition) Addison-Wesley Longman Inc.
- INGRAHAM J.L. & INGRAHAM C.A. (1995) Introduction to Microbiology Wadsworth Publishing Co..
- PERRY J.J. & STALEY J.T. (1997) Microbiology Dynamics and Diversity Saunders College Publishing
- TORTORA, G.J., FUNKE, B.R. & CASE C.L. (1995) Microbiology An Introduction Benjamin Cummings Publishers
- VOLK W.A. & BROWN J.C. (1997) Basic Microbiology (8th Edition)
  Benjamin Cummings Publishers

#### Text-books on World Wide Web:

'Medical Microbiology' (4th Edition) Editor: Samuel Baron http://129.109.136.65/microbook/toc.html

\*Microbiology 101 Internet Text' (Washington State University)
http://www.wsu.edu/~hurlbert-pages-101hmpg.html

#### Other Available Resources:

#### MI 2650 web page at GPRC:

http://www.gprc.ab.ca/courses\_and\_programs/biology/mi2650nf.htm

#### MI 265 web page at University of Alberta:

http://gause.biology.ualberta.ca/courses.hp/micrb.htm

#### Requirements\_\_\_\_

Lab. Reports			į		4	15%
Quizzes						
Mid-term Exam	ě	8	-			20%
Final Lab Exam						
						35%
TOTAL					1	00%

Each student should maintain a card file on the significant bacteria mentioned in class. Keep a record of: Genus and species; cell morphology; Gram stain reaction; habitat; 4 or 5 interesting facts about the organism's growth, metabolism, pathogenicity, use in industry, etc. Do not keep records of taxonomic tests. Information can be obtained from text-books, lectures, "Bergey's Manual of Determinative Bacteriology", the Internet, or other sources. A question related to this information will appear on both the Mid-term and Final Exams.

In order to successfully complete MI 2650, students must attend ALL laboratory sessions and achieve a mean score of 50% on the Lab Reports, Lab Quizzes and Final Lab Exam. All assignments MUST be handed in by the time and date specified. Late reports will not be marked!

Many of the Laboratory exercises require that students perform some of the procedures at times other than the scheduled lab period. To do this, prior arrangements must be made with Mr. Rick Scott, the Biology Lab Technologist. In case of injury, it is preferable that students work with at least one partner when coming into the laboratory outside of scheduled times.

Quizzes in both class and laboratory sessions may be given without any advanced notice to students.

Since participation in lectures, and completion of assignments are important components of this course, students will serve their best interests by regular attendance. Those who chose not to attend must assume whatever risks are involved. In this regard, your attention is directed to the Academic Guidelines of Grande Prairie Regional College.

#### MI 2650 TOPIC OUTLINE

TOPIC	READINGS
Introduction to the course	2-12, 15-17, 26-28, 422-424
Functional morphology:	
Definitions and descriptions of microbes	50-53, 57-60, 77-79, 99-100, 237-238, 285-287, 432-442, 449-51, 724-726, 729, 733, 735-736
Cell structure and function	60-78, 91-95
Structural features in beneficial and harmful bacteria	85-87, 502-506, 645-647, 774-782, 784-788, 791-793, 923-929, 940-943, 974-976, 986-987, 988-989, 20-24
Motility and chemotaxis	79-85, 231-233, 495, 567-538
Microbial diversity and environments:	
Major nutritional types based on carbon and energy source	103-106, 642-650
Review of aerobic and anaerobic respiration	108-129, 601-603, 605-608,
Fermentation	118-121, 406-407, 477-482, 620-626, 504-506,
Photosynthesis	574-591
Practical examples of diverse nutritional types:	
methanogenesis	416-420, 553-556, 613-617, 681-685
bioleaching and bioremediation	631-634, 691-694, 696-703
extremophiles	670-675

#### TOPIC (continued) READINGS Microbial growth: Growth in relation to oxygen (use and toxicity) 158-162 Effects of temperature, nutrient levels and growth conditions 88, 147-158 Exponential growth curve 139-147 Prediction of growth rate and cell yield 136-139 Control of growth. heat, filters, etc. 742-749 chemicals (heavy metals, antibiotics) 387-389, 392-399, 749-762 Resistance to chemical agents (especially antibiotics) 765-772 Development of new antibacterial agents Sensory systems and intercellular communication: Transcriptional control systems in bacteria, sigma factors 191-194 Global regulation and quorum sensing 226-233 Nitrogen cycle and regulation 634-639, 685-686 Plant-microbe interactions: Rhizobium spp 709-717 Agrobacterium spp. 706-708 Biotechnology: Recombinant DNA technology 368-374, 378-382 Agrobacterium Ti plasmid and transgenic plants 374-376

#### MI 2650 LABORATORY SCHEDULE WINTER 1999-2000

DATE	EXERCISE	STEPS	TOPICS
January 14	E	A1-4 B1	Isolation of component bacteria from a mixture.  Gram stain of the bacteria in the mixture
	2	A1-4 B1-2 C1 D1	Standard Method Plate Count Isolation and identification of Escherichia coli Enrichment of Streptococcus faecalis Enrichment of Staphylococcus aureus
January 17	1		Check TCS plates and isolate pure cultures
	2	A5 B3-5 C2-3 D2-3	Read plate counts  Isolation and identification of E. coli (contd.)  Enrichment of S. faecalis (contd.)  Enrichment of S. aureus (contd.)
January 21 1	1		Perform Oxidase test on each pure culture Identification of Gram negative enteric bacteria (API 20E strip
	2	B6 C4-6 D4-5	Gram stain of pure E. coli  Gram stain and Catalase test on presumed faecal streptococci  Gram stain and Coagulase test on β-hemalytic bacterium
	3	1-9 D1-4 E1-2	Preparation of yogurt  Examination of commercial yogurt  Examination of culture after incubation at 46-48°C
January 24	1		Read API 20E strip results
	2	C7	Check BHI tubes for growth
January 28			HAND IN REPORT FROM EXERCISE 1
	3	10-13	Examination of prepared yogurt
	4		Identification of unknown bacteria
	5	A1-2	Preparation of enzyme from Trichoderma cultures
January 31	4		Obtain results from: MacConkey's Medium  EMB agar  Blood haemolysis
February 4			HAND IN REPORT FROM EXERCISE 3
	4	A10-11 B7	
	5	A3 B1-6 C1-3	Filtration of Test Enzyme Enzyme assay Measurement of reducing sugar

Laboratory Schedule (continued)

February 11			HAND IN REPORT FROM EXERCISE 2				
	7	all	Regulation of the law operon				
February 18	NO LAB		HAND IN REPORT FROM EXERCISE 4				
February 25	NO LAB		WINTER BREAK				
March 3	6	A1-7 B1-4	HAND IN REPORT FROM EXERCISE 5 Nitrogen-fixation				
		C1-9	Denitrification				
	8	1-2	Production of penicillin				
March 10			HAND IN REPORT FROM EXERCISE 7				
	6	B5-10 C10	Ammonification (contd.) Denitrification (contd.)				
	8	A3-10 B1-5	Quantification of penicillin Activity spectra				
	9	A1-2 B1-8	Preparation of competent E. coli cells Transformation of E. coli				
March 13	8		Determine Optical Density of bioassay tubes Measure Zones of Inhibition				
	9	C1-3	Selection of transformed E. coli				
March 17			HAND IN REPORT FROM EXERCISE 6				
3/	- 9	D1-2 E1-3	Aldehyde detection Ethanol production by E. coli				
March 20	9	E4-5	Ethanol production by E. coli (contd.)				
March 24			HAND IN REPORT FROM EXERCISE 8				
	9	F1 G1-19	pH of fermentation broth Quantitiation of ethanol: dichromate reduction				
March 31			HAND IN REPORT FROM EXERCISE 9  LAB EXAM (written)				